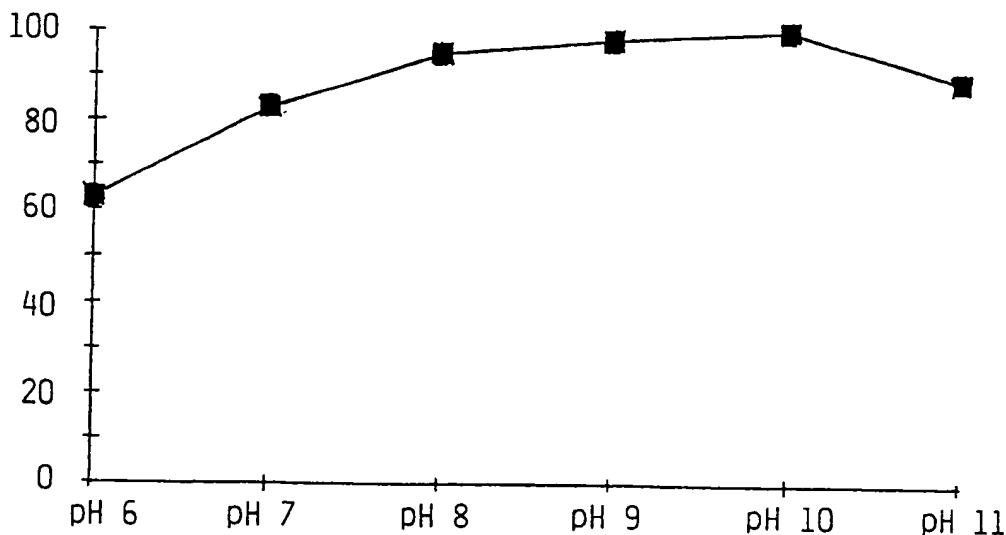


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(21) International Application Number: PCT/DK92/00104 (22) International Filing Date: 3 April 1992 (03.04.92) (30) Priority data: 584/91 3 April 1991 (03.04.91) DK (71) Applicant (for all designated States except US): NOVO NORDISK A/S [DK/DK]; Novo Allé, DK-2880 Bagsvaerd (DK). (72) Inventors; and (75) Inventors/Applicants (for US only): OUTTRUP, Helle [DK/DK]; Syvendehusvej 46, DK-2750 Ballerup (DK). DAMBMANN, Claus [DK/DK]; Hoeje Gladsaxe 61, 7.t.h., DK-2860 Soeborg (DK). LINDEGAARD, Poul [DK/DK]; Oester Soegade 52, DK-2100 Copenhagen Ø (DK).		(74) Common Representative: NOVO NORDISK A/S; Patent Department, Novo Allé, Dk-2880 Bagsværd (DK). (81) Designated States: AT (European patent), BE (European patent), CH (European patent), DE (European patent), DK (European patent), ES (European patent), FI, FR (European patent), GB (European patent), GR (European patent), IT (European patent), JP, KR, LU (European patent), MC (European patent), NL (European patent), SE (European patent), US. Published <i>With international search report.</i>

(54) Title: NOVEL PROTEASES**(57) Abstract**

This invention is in the field of detergent proteases derived from strains of a new *Bacillus* sp. More specifically, the invention is directed towards a protease derived from a strain of *Bacillus* sp. TY 145. Moreover, the invention is directed towards a process for the preparation of the protease, the use of the protease as detergent enzyme, and detergent compositions comprising the protease of the invention.

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NOVEL PROTEASES

TECHNICAL FIELD

This invention is in the field of detergent proteases derived from strains of *Bacillus* sp. More specifically, the invention is directed towards a novel alkaline protease derived from a strain of *Bacillus* sp. TY145. Moreover, the invention is directed towards a process for the preparation of the protease, the use of the protease as detergent enzyme, and detergent compositions comprising the protease of the invention.

BACKGROUND ART

Detergent enzymes have been marketed for more than 20 years and are now well established as normal detergent ingredients in both powder and liquid detergents all over the world. With the trend towards lower temperature washing, detergent enzyme consumption has increased during late years. Enzymes used in washing formulations comprise proteases, lipases, amylases, cellulases, as well as other enzymes, or mixtures hereof. Commercially most important are proteases.

Detergent proteases have been developed by isolation of proteases found in nature followed by testing in detergent formulations. Most detergent proteases are obtained from members of the genus *Bacillus*. Currently new types of proteases enter the market, offering the possibility of giving a better cost/performance ratio at various specified conditions.

Examples of commercial protease products are ALCALASE™, ESPERASE™ and SAVINASE™, all supplied by Novo Nordisk A/S, Denmark. These and similar enzyme products from other commercial sources are active in detergent solutions, i.e. at pH values in the range of from 8 to 11 and in the presence of sequestering agents, surfactants and bleaching agents such as sodium borate. The ALCALASE™ protease is produced by strains of the species *Bacillus licheniformis*. The ESPERASE™ and SAVINASE™ proteases are obtained by cultivation of strains of alkalophilic *Bacilli*.

It is an object within the present invention to provide novel detergent proteases with improved cost/performance ratio.

SUMMARY OF THE INVENTION

According to the present invention there are provided novel detergent
5 proteases.

In its first aspect, the invention provides a protease having an apparent molecular weight of 38 kD, a pI around 8.8, pH optimum in the range of from pH 8 to 11 (at 25°C), temperature optimum in the range of from 45 to 55 °C (at pH 9.5), and immunochemical properties identical or partially identical to those of a protease
10 derived from *Bacillus sp.* TY145, NCIMB No. 40339. In a more specific aspect, the protease is obtainable from a strain of *Bacillus sp.* TY145. In a yet more specific aspect, the protease is obtainable from *Bacillus sp.* TY145, NCIMB No. 40339, or a mutant or a variant thereof.

In another aspect, the invention provides an isolated biologically pure
15 culture of a strain of *Bacillus sp.* TY145. In a more specific aspect, a strain of *Bacillus sp.* TY145, NCIMB No. 40339, or a mutant or a variant thereof, is provided.

In a third aspect, the invention provides a process for the preparation of the protease, which process comprises cultivation of a protease producing strain of *Bacillus sp.* TY145 in a suitable nutrient medium, containing carbon and nitrogen
20 sources and inorganic salts, followed by recovery of the desired enzyme. In a more specific aspect, *Bacillus sp.* TY145, NCIMB No. 40339, or a mutant or a variant thereof, is cultivated.

In a fourth aspect, the use of the enzyme as detergent enzyme is claimed. In a more specific aspect, the invention provides a detergent composition
25 and a detergent additive comprising the protease.

BRIEF DESCRIPTION OF DRAWINGS

The present invention is further illustrated by reference to the accompanying drawings, in which:

Fig. 1 shows the relation between temperature and the proteolytic activity of an enzyme according to the invention (the enzyme preparation obtained according to Ex. 1, with casein as substrate and at pH 9.5); and

Fig. 2 shows the relation between pH and the proteolytic activity of an enzyme according to the invention (the enzyme preparation obtained according to Ex. 1, with casein as substrate and at 25°C).

DETAILED DISCLOSURE OF THE INVENTION

The microorganism

The novel microorganism of the invention, able to produce an enzyme of the invention, was isolated from a sample of Antarctic soil. *Bacillus sp.* TY145 has been deposited according to the Budapest Treaty at NCIMB, under No. 40339.

The microorganism of this invention is an aerobic, spore forming bacterium belonging to the genus *Bacillus*. Morphologically it can be described as motile rods with a diameter of 0.7 - 0.9 micron, and a length of 2 - 3 micron. The spores are round to ellipsoid, not swelling the sporangium, central to subterminal. Optimal temperature for growth is within 30 - 37°C, and optimal pH for growth is within 7 - 9, no growth at pH 9.7, and no growth at 50°C. The microorganism forms dry, white colonies with hairy outgrowths on nutrient agar slants, and no diffusion of pigment into the agar is observed.

20 Cultivation of the microorganism

The microorganism of the invention can be cultivated under aerobic conditions in a nutrient medium containing assimilable carbon and nitrogen together with other essential nutrients, the medium being composed in accordance with the principles of the known art.

25 Suitable carbon sources are carbohydrates such as sucrose, glucose and starch, or carbohydrate containing materials such as cereal grain, malt, rice and sorghum. The carbohydrate concentration incorporated in the medium may vary widely, e.g. up to 25% and down to 1 - 5%, but usually 8 - 10% will be suitable, the percentages being calculated as equivalents of glucose.

The nitrogen source in the nutrient medium may be of inorganic and/or organic nature. Suitable inorganic nitrogen sources are nitrates and ammonium salts. Among the organic nitrogen sources quite a number are used regularly in fermentation processes involving the cultivation of bacteria. Illustrative examples are soybean meal, cotton seed meal, peanut meal, casein, corn, corn steep liquor, yeast extract, 5 urea and albumin. In addition, the nutrient medium should also contain usual trace substances.

The novel *Bacillus* species of this invention are slightly alkalophilic. Therefore, the cultivation is preferably conducted at slightly alkaline pH values, which 10 can be obtained by addition of suitable buffers such as sodium bicarbonate, pH 9.0, after sterilization of the growth medium. For cultivation in tank fermentors it is necessary to use artificial aeration. The rate of aeration is similar to that used in conventional tank fermentation.

After fermentation, liquid enzyme concentrates may be produced by 15 removal of coarse material from the broth or, if desired, concentration of the broth by evaporation at low temperature or by reverse osmosis. Finally, preservatives may be added to the concentrate.

Solid enzyme preparations may be prepared from the purified and/or concentrated broth by precipitation with salts, such as Na_2SO_4 or water-miscible 20 solvents, such as ethanol or acetone, removal of the water in the broth by suitable drying methods such as spray-drying may also be employed.

Assay for proteolytic activity

The proteolytic activity is determined with casein as substrate. One Casein Protease Unit (CPU) is defined as the amount of enzyme liberating 1 mM of 25 primary amino groups (determined by comparison with a serine standard) per minute under standard conditions, i.e. incubation for 30 minutes at 25°C and pH 9.5. A folder AF 228, describing the analytical method, is available upon request to Novo Nordisk A/S, Denmark, which folder is hereby included by reference.

The enzyme

The enzyme of the invention is a novel detergent protease. It is an alkaline protease, obtainable by cultivation of a microorganism of the invention, preferably *Bacillus sp.* TY145, NCIMB No. 40339, or a mutant or a variant thereof, in a suitable nutrient medium, containing carbon and nitrogen sources and inorganic salts. The enzyme can also be obtained by recombinant DNA-technology.

The protease of the invention can be described by the following characteristics.

Physical-chemical properties

10 A molecular weight of 38 kD, determined by SDS-PAGE. A pI of 8.8 determined by isoelectric focusing on LKB Ampholine® PAG plates. The protease activity is inhibited by PMSF, α -1-antitrypsin and Turkey-egg-white proteinase inhibitor. EDTA and soybean-protein inhibitor do not influence the protease activity.

The temperature activity relationship was determined with casein as 15 substrate. The assay for proteolytic activity described previously was used with the modification that the incubation temperature was varied in the interval of from 15 to 70°C. The result is shown in Fig. 1. The enzyme possesses proteolytic activity from temperatures below 15°C to above 70°C, and a temperature optimum within the range 45 to 55°C, around 50°C.

20 The dependence of activity on pH was determined by the same procedure, using buffers adjusted to predetermined pH values in the pH range of from 6 to 11. The result is shown in Fig. 2. The enzyme possesses proteolytic activity at pH values below 6 to above 11, with a pH optimum in the range pH 8 to pH 11, around pH 10.

25 The protease of the invention is stable for 60 minutes at 40°C under washing conditions, in European type and American type detergents.

Immunochemical properties

The immunochemical properties can be determined immunologically by cross-reaction identity tests. The identity tests can be performed by the well-known Ouchterlony double immunodiffusion procedure or by tandem crossed 30

immuno-electrophoresis according to *Axelsen, N.H.*; Handbook of Immuno-precipitation-in-Gel Techniques; Blackwell Scientific Publications (1983), chapters 5 and 14. The terms "antigenic identity" and "partial antigenic identity" are described in the same book, chapters 5, 19 and 20.

5 Monospecific antiserum was generated according to the above mentioned method by immunizing rabbits with the purified protease of the invention. The immunogen was mixed with Freund's adjuvant and injected subcutaneously into rabbits every second week. Antiserum was obtained after a total immunization period of 8 weeks, and immunoglobulin was prepared therefrom as described by N. H.
10 *Axelsen, supra*.

Ouchterlony double immunodiffusion tests showed no cross reaction between the protease of the invention and the known alkaline serine proteases ALCALASE™, SAVINASE™, ESPERASE™, subtilisin BPN' and KAZUSASE™.

Detergent compositions

15 The detergent composition of the invention may comprise one or more surfactants, which may be of an anionic, non-ionic, cat-ionic, amphoteric or zwitter-ionic type, or a mixture of these. Typical examples of anionic surfactants are linear alkyl benzene sulfonates (LAS), alkyl sulfates (AS), alpha olefin sulfonates (AOS), alcohol ethoxy sulfates (AES) and alkali metal salts of natural fatty acids. Examples
20 of non-ionic surfactants are alkyl polyethylene glycol ethers, nonylphenol polyethylene glycol ethers, fatty acids esters of sucrose and glucose, and esters of polyethoxylated alkyl glucoside.

The detergent composition of the invention may also contain other detergent ingredients known in the art such as builders, bleaching agents, bleach
25 activators, anti-corrosion agents, sequestering agents, anti soil-redeposition agents, perfumes, stabilizers for the enzymes and bleaching agents, formulations aids, optical brighteners, foam boosters, chelating agents, fillers, fabric softeners, etc. The detergent composition of the invention may be formulated substantially as described in J. Falbe [*Falbe, J.*; Surfactants in Consumer Products. Theory, Technology and
30 Application; Springer Verlag 1987, *vide* in particular the section entitled "Frame formulations for liquid/powder heavy-duty detergents"].

It is at present contemplated that the detergent composition of the invention may contain the enzyme preparation in an amount corresponding to 0.0005-0.5 CPU of the proteolytic enzyme per litre of washing liquor.

The detergent compositions of the invention can be formulated in any
5 convenient form, such as powders, liquids, etc.

The detergent composition of the invention may advantageously include one or more other enzymes, e.g. lipases, amylases, cellulases, oxidases, and/or peroxidases, conventionally included in detergent compositions.

The protease of the invention may be included in a detergent
10 composition by adding separate additives containing the detergent protease, or by adding a combined additive comprising different detergent enzymes.

The additive of the invention can be formulated e.g. as granulates, liquids, slurries, etc. Preferred detergent additive formulations are non-dusting granulates, liquids, in particular stabilized liquids, slurries, or protected enzymes.
15 Dust free granulates may be produced e.g. according to GB Patent Publication No. 1,362,365 or US Patent No. 4,106,991, and may optionally be coated by methods known in the art. The detergent enzymes may be mixed before or after granulation. Liquid enzyme preparations may, for instance, be stabilized by adding a polyol such as e.g. propylene glycol, a sugar or sugar alcohol, lactic acid or boric acid,
20 according to established methods. Other enzyme stabilizers are well known in the art. Protected enzymes may be prepared according to the method disclosed in EP Patent Publication No. 238,216.

In usefull embodiments the protease of the invention may be incorporated in detergent formulations according to e.g. EP Patent Publication Nos.
25 342,177; 368,575; 378,261; and 378,262.

The invention is further illustrated in the following examples, which are not intended to be in any way limiting to the scope of the invention as claimed.

EXAMPLE 1

Cultivation Example

Bacillus sp. TY145 was cultivated at 25°C on a rotary shaking table (300 r.p.m.) in 500 ml baffled Erlenmeyer flasks containing 100 ml of medium of the following composition (per litre):

Potato starch	100 g
Ground barley	50 g
Soybean flour	20 g
Na ₂ HPO ₄ x 12 H ₂ O	9 g
Pluronic®	0.1 g
Sodium caseinate	10 g

The starch in the medium is liquified with α -amylase and the medium is sterilized by heating at 120°C for 45 minutes.

After sterilization the pH of the medium is adjusted to 9.0 by addition of 10 ml of a 1 M solution of sodium bicarbonate.

After 5 days of incubation the proteolytic activity of the culture was determined using the method described above.

After cultivation, the enzyme activity of the broth was 10 CPU/l.

After separation of the solid material the protease was purified by a conventional chromatographic method.

Yield from 1 l of culture broth was 50 ml with 57 CPU/l. Purity was more than 90% as judged by SDS-PAGE.

The characteristics of the preparation prepared in accordance with this Example have been referred to earlier in this specification, and reference is made hereto.

EXAMPLE 2

Wash performance

The wash performance tests were accomplished on grass soiling on cotton, in a model wash system at 20°C, isothermically for 10 minutes.

5 2.0 g/l of a commercial American type liquid detergent were used in the test. The detergent did not contain any enzymes prior to the addition of the protease of the invention. The detergent was dissolved in approx. 6°dH (German Hardness) water, and the pH was measured to approx. 8. The textile/wash liquor ratio was approximately 6 g textile per litre of detergent solution. The enzyme preparation
10 according to Example 1 was used at enzyme protein concentrations of 0.01; 0.04; 0.08; 0.16; and 0.5 CPU/l.

Subsequent to washing, the fabric was rinsed in running tap water for 25 minutes and air-dried. The protease performance was determined by the change (ΔR) of the remission (%R) at 460 nm measured on a Datacolor Elrephometer 2000,
15 ΔR being the remission after wash with protease added minus the remission after wash with no protease added.

The test results are shown in Table 1.

Table 1

	Enzyme Concentration	delta R
20		
	0.01 CPU/l	4
	0.04 CPU/l	6.7
	0.08 CPU/l	8.7
25	0.16 CPU/l	9.7
	0.50 CPU/l	11.2

The differential remission values show that the protease of the invention possesses good washability.

EXAMPLE 3Stability in detergents

The stability of an enzyme of the invention (the enzyme preparation obtained according to Ex. 1) was tested in the presence of detergents. The 5 detergents used in this test were an American type powder detergent and an American type liquid detergent.

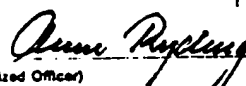
The residual activity was determined after 60 minutes at 40°C. Enzyme dosage was 0.3 CPU/l.

			Residual activity
10	Powder detergent:	0.9 g/l	100%
	Liquid detergent:	2.0 g/l	95%

This experiment shows that the protease is stable in detergents under 15 washing conditions.

International Application No: PCT/

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MICROORGANISMS	
Optional Sheet in connection with the microorganism referred to on page <u>1</u> , line <u>3</u> of the description ¹	
A. IDENTIFICATION OF DEPOSIT ¹	
Further deposits are identified on an additional sheet <input type="checkbox"/> ²	
Name of depositary institution ⁴ NATIONAL COLLECTIONS OF INDUSTRIAL & MARINE BACTERIA LTD.	
Address of depositary institution (including postal code and country) ⁴ 23 St. Machar Drive, Aberdeen AB2 1RY, Scotland	
Date of deposit ⁵ 3 December 1990	Accession Number ⁶ NCIMB 40339
B. ADDITIONAL INDICATIONS ⁷ (leave blank if not applicable). This information is continued on a separate attached sheet <input type="checkbox"/>	
In respect of those designations in which a European patent is sought, a sample of the deposited micro-organism will be made available only by the issue of such a sample to an expert nominated by the person requesting the sample (Rule 28(4) EPC) until the publication of the mention of the grant of the European patent or until the date on which the application has been refused or is deemed to be withdrawn.	
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 (Authorized Officer)	
<input type="checkbox"/> The date of receipt (from the applicant) by the International Bureau ¹⁰	
was	_____ (Authorized Officer)

CLAIMS

1. A protease characterized by having the following properties:
 - (a) an apparent molecular weight of 38 kD;
 - (b) pI around 8.8;
 - 5 (c) pH optimum in the range of from pH 8 to 11 (at 25°C);
 - (d) temperature optimum in the range of from 45 to 55°C (at pH 9.5);
 - (e) immunochemical properties identical or partially identical to those of a protease derived from *Bacillus sp.* TY145, NCIMB No. 40339.
2. A protease according to claim 1, the protease being obtainable from
10 a strain of *Bacillus sp.* TY145.
3. The protease of claim 2, being obtainable from *Bacillus sp.* TY145, NCIMB No. 40339, or a mutant or a variant thereof.
4. An isolated biologically pure culture of a strain of *Bacillus sp.* TY145.
5. A culture according to claim 4, the strain being *Bacillus sp.* TY145,
15 NCIMB No. 40339, or a mutant or a variant thereof.
6. A process for the preparation of a protease according to any of claims 1-3, which process comprises cultivation of a protease producing strain of *Bacillus sp.* TY145 in a suitable nutrient medium, containing carbon and nitrogen sources and inorganic salts, followed by recovery of the desired enzyme.
- 20 7. A process according to claim 6, in which *Bacillus sp.* TY145, NCIMB No. 40339, or a mutant or a variant thereof, is cultivated.
8. The use of a protease according to any of claims 1-3 as a detergent enzyme.

9. A detergent composition comprising a protease according to any of claims 1-3.

10. A detergent composition according to claim 9, which further comprises one or more other enzymes, in particular amylases, lipases, cellulases, oxidases, and/or peroxidases.

11. A detergent composition comprising a protease according to any of claims 1-3, provided in the form of a non-dusting granulate, a liquid, in particular a stabilized liquid, a slurry, or a protected enzyme.

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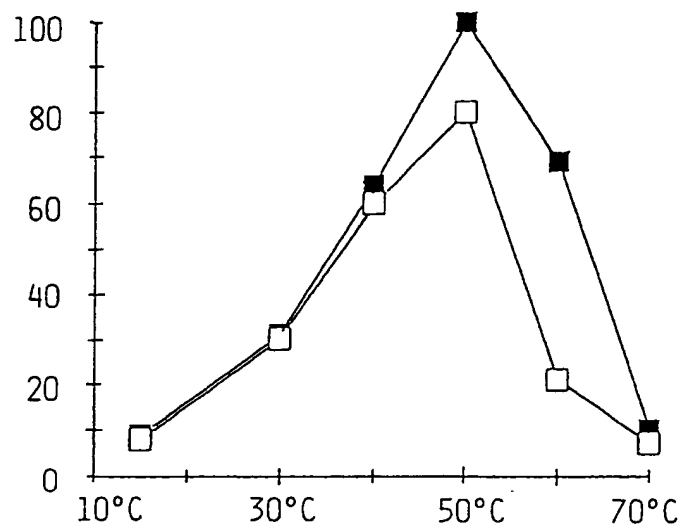


Fig. 1

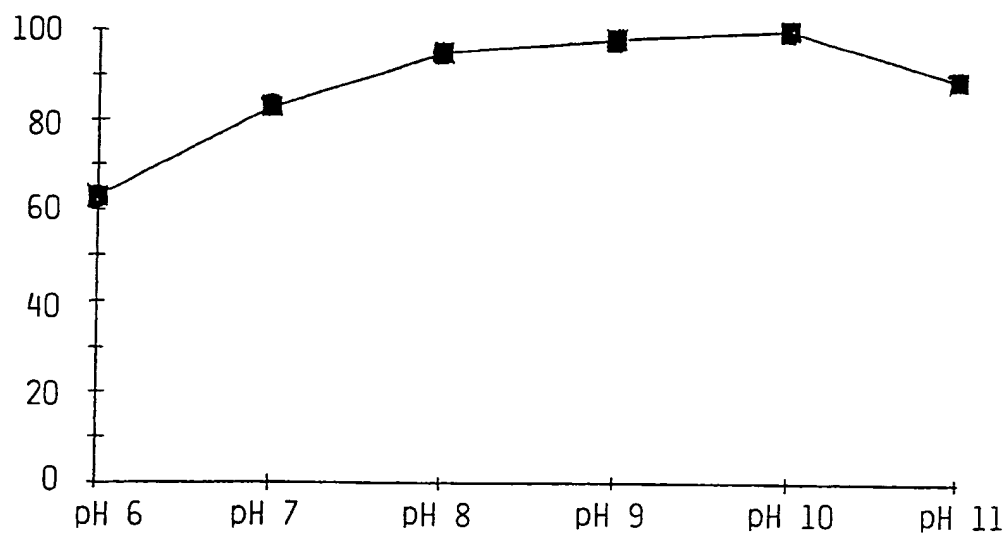


Fig. 2

INTERNATIONAL SEARCH REPORT

International Application No PCT/DK 92/00104

I. CLASSIFICATION OF SUBJECT MATTER (If several classification symbols apply, indicate all) ⁶ According to International Patent Classification (IPC) or to both National Classification and IPC IPC5: C 12 N 9/54, C 11 D 3/386											
II. FIELDS SEARCHED <div style="text-align: right; font-size: small;">Minimum Documentation Searched⁷</div> <table style="width: 100%; border: none;"> <tr> <td style="width: 25%; border: none;">Classification System</td> <td style="border: none;">Classification Symbols</td> </tr> <tr> <td style="border: 1px solid black; padding: 5px;">IPC5</td> <td style="border: 1px solid black; padding: 5px;">C 12 N; C 11 D</td> </tr> </table> <div style="text-align: center; font-size: x-small; margin-top: 5px;">Documentation Searched other than Minimum Documentation to the Extent that such Documents are Included in Fields Searched⁸</div> <p style="margin-top: 10px;">SE,DK,FI,NO classes as above</p>			Classification System	Classification Symbols	IPC5	C 12 N; C 11 D					
Classification System	Classification Symbols										
IPC5	C 12 N; C 11 D										
III. DOCUMENTS CONSIDERED TO BE RELEVANT⁹ <table border="1" style="width: 100%; border-collapse: collapse;"> <thead> <tr> <th style="width: 10%; font-size: x-small;">Category *</th> <th style="width: 60%; font-size: x-small;">Citation of Document,¹¹ with indication, where appropriate, of the relevant passages¹²</th> <th style="width: 30%; font-size: x-small;">Relevant to Claim No.¹³</th> </tr> </thead> <tbody> <tr> <td style="text-align: center; vertical-align: top;">X</td> <td style="vertical-align: top;"> <p>Dialog Information Services, File 357, Biotechnology Abstract, 82-92, Dialog accession no. 092501, DBA accession no. 89-10492, LION: "Production of a novel alkaline protease-L - isolation from Bacillus sp. and characterization", Patent number: JP 1101884, Patent Date: 890419.</p> <p style="text-align: center;">---</p> </td> <td style="text-align: center; vertical-align: top;">1-11</td> </tr> <tr> <td style="text-align: center; vertical-align: top;">A</td> <td style="vertical-align: top;"> <p>EP, A2, 0204342 (LION CORPORATION) 10 December 1986, see the whole document</p> <p style="text-align: center;">-----</p> </td> <td style="text-align: center; vertical-align: top;">1-11</td> </tr> </tbody> </table>			Category *	Citation of Document, ¹¹ with indication, where appropriate, of the relevant passages ¹²	Relevant to Claim No. ¹³	X	<p>Dialog Information Services, File 357, Biotechnology Abstract, 82-92, Dialog accession no. 092501, DBA accession no. 89-10492, LION: "Production of a novel alkaline protease-L - isolation from Bacillus sp. and characterization", Patent number: JP 1101884, Patent Date: 890419.</p> <p style="text-align: center;">---</p>	1-11	A	<p>EP, A2, 0204342 (LION CORPORATION) 10 December 1986, see the whole document</p> <p style="text-align: center;">-----</p>	1-11
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<div style="font-size: x-small;"> <p>* Special categories of cited documents: ¹⁰</p> <p>"A" document defining the general state of the art which is not considered to be of particular relevance</p> <p>"E" earlier document but published on or after the international filing date</p> <p>"L" document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another citation or other special reason (as specified)</p> <p>"O" document referring to an oral disclosure, use, exhibition or other means</p> <p>"P" document published prior to the international filing date but later than the priority date claimed</p> <p>"T" later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention</p> <p>"X" document of particular relevance, the claimed invention cannot be considered novel or cannot be considered to involve an inventive step</p> <p>"Y" document of particular relevance, the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art.</p> <p>"&" document member of the same patent family</p> </div>											
IV. CERTIFICATION <table style="width: 100%; border: none;"> <tr> <td style="width: 50%; border: none; vertical-align: top;"> <div style="border: 1px solid black; padding: 5px; margin-bottom: 5px;">Date of the Actual Completion of the International Search</div> <div style="border: 1px solid black; padding: 5px; margin-bottom: 5px;">2nd July 1992</div> <div style="border: 1px solid black; padding: 5px;">International Searching Authority</div> <div style="text-align: center; margin-top: 10px;">SWEDISH PATENT OFFICE</div> </td> <td style="width: 50%; border: none; vertical-align: top;"> <div style="border: 1px solid black; padding: 5px; margin-bottom: 5px;">Date of Mailing of this International Search Report</div> <div style="border: 1px solid black; padding: 5px; margin-bottom: 5px;">1992 -07- 07</div> <div style="border: 1px solid black; padding: 5px;">Signature of Authorized Officer</div> <div style="text-align: center; margin-top: 5px;"> <i>Carolina Palmcrantz</i> Carolina Palmcrantz </div> </td> </tr> </table>			<div style="border: 1px solid black; padding: 5px; margin-bottom: 5px;">Date of the Actual Completion of the International Search</div> <div style="border: 1px solid black; padding: 5px; margin-bottom: 5px;">2nd July 1992</div> <div style="border: 1px solid black; padding: 5px;">International Searching Authority</div> <div style="text-align: center; margin-top: 10px;">SWEDISH PATENT OFFICE</div>	<div style="border: 1px solid black; padding: 5px; margin-bottom: 5px;">Date of Mailing of this International Search Report</div> <div style="border: 1px solid black; padding: 5px; margin-bottom: 5px;">1992 -07- 07</div> <div style="border: 1px solid black; padding: 5px;">Signature of Authorized Officer</div> <div style="text-align: center; margin-top: 5px;"> <i>Carolina Palmcrantz</i> Carolina Palmcrantz </div>							
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ANNEX TO THE INTERNATIONAL SEARCH REPORT ON INTERNATIONAL PATENT APPLICATION NO. PCT/DK 92/00104

This annex lists the patent family members relating to the patent documents cited in the above-mentioned international search report. The members are as contained in the Swedish Patent Office EDP file on 29/05/92. The Swedish Patent Office is in no way liable for these particulars which are merely given for the purpose of information.

Patent document cited in search report	Publication date	Patent family member(s)	Publication date
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		JP-A- 61280268	86-12-10
		US-A- 4797362	89-01-10
		JP-B- 3000997	91-01-09
		JP-A- 61280278	86-12-10
		JP-B- 2055034	90-11-26
		JP-A- 62146594	87-06-30